

学位論文の内容の要約

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学位論文題目	Analysis of Susceptibility-determining Receptors of <i>Bombyx mori</i> for <i>Bacillus thuringiensis</i> Cry toxins

【論文の内容の要約】

Field-evolved resistance of insect pests to *Bacillus thuringiensis* (Bt) toxins (Cry toxins) is a threat to the efficacy of Bt-based bio-insecticides and transgenic crops. Recent reports have suggested that ATP-binding cassette transporters and cadherin-like receptor play important roles in conferring susceptibility to Cry toxins. However, the receptors involved in Bt susceptibility in each insect remain unclear.

In the Chapter 1, to determine the receptors that are involved in the susceptibility of *Bombyx mori* to Cry1 toxins (1Ab, 1Ac and 1Fa), I conducted diet overlay bioassay using *B. mori* strains disrupted with one or two receptor (s) among ATP-binding cassette transporter family C2 (BmABCC2), ATP-binding cassette transporter family C3 (BmABCC3), and cadherin-like receptor (BtR175) generated by transcription activator-like effector nuclease (TALEN)-mediated gene editing. The single-knockout strains for *BmABCC2* showed resistance to Cry1Ab and Cry1Ac, whereas only strains with double knockout of *BmABCC2* and *BmABCC3* exhibited high resistance to Cry1Fa. Progeny populations generated from the crossing of heterozygotes for BtR175 knockout allele included 25% theoretical homozygotes for the BtR175 knockout allele and they showed resistance to Cry1Ab and Cry1Ac. Then, through a cell swelling assay using Sf9 cells ectopically expressing the receptor, I analyzed the mechanisms underlying the different contributions of *BmABCC2*, *BmABCC3*, and BtR175 to larval susceptibility. The receptor activity of *BmABCC2* for Cry1Ab and Cry1Ac was far higher than that of *BmABCC3*, and BtR175 synergistically enhanced the receptor activity of *BmABCC2*. This result well explained the important involvement of *BmABCC2* and BtR175 in the larval susceptibility to Cry1A toxins. By contrast, the receptor activities of *BmABCC2* and *BmABCC3* for Cry1Fa were observed at a similar level and synergistic effect of BtR175 was small. This finding explains the equal importance of *BmABCC2* and *BmABCC3* and very small contribution of BtR175 on larval susceptibility to Cry1Fa. Thus, I demonstrated the different importance of *BmABCC2*, *BmABCC3*, and BtR175 to various Cry1 toxins as susceptibility-determining factors in *B. mori* larvae and the underlying basis for the observed differences. Furthermore, a weak correlation was indicated between the binding affinity and receptor activities of *BmABCC2* and *BmABCC3* to Cry1 toxins.

In the Chapter 2, I investigated whether *BmABCC1* and *BmABCC4* could be functional receptor in *B. mori* to several Cry toxins, Cry1Aa, Cry1Da, Cry8Ca and Cry9Aa. Firstly, I did a cell swelling assay using *BmABCC1* and *BmABCC4* expressing Sf9 cells to explore whether they do have a receptor function to several Cry toxins. The *BmABCC1* showed receptor activity to Cry9Aa, and *BmABCC4* showed receptor activity to Cry1Aa, Cry1Da, Cry8Ca. However, *BmABCC1* knockout *B. mori* larvae did not show resistance to Cry9Aa and *BmABCC4* knockout *B. mori* larvae did not show resistance to Cry1Aa, Cry1Da, Cry8Ca. These results suggest that *BmABCC1* and *BmABCC4* can function as low performance receptors to Cry1Aa, Cry1Da, Cry8Ca and Cry9Aa toxins because *BmABCC1* and *BmABCC4* did not contribute to the determination of susceptibility of *B. mori* larvae to those Cry toxins and a low binding affinity of *BmABCC1* with Cry9Aa, and *BmABCC4* with Cry1Aa, Cry1Da and Cry8Ca was also observed in SPR analysis.